

# Conservation implications of the evolutionary history and genetic diversity hotspots of the snowshoe hare

ELLEN CHENG,\* KAREN E. HODGES,† JOSÉ MELO-FERREIRA,‡ PAULO C. ALVES\*§ and L. SCOTT MILLS\*¶

\*Wildlife Biology Program, Department of Ecosystem and Conservation Sciences, University of Montana, 32 Campus Drive, Missoula, MT 59812, USA, †Department of Biology, University of British Columbia Okanagan, 3333 University Way, Kelowna, BC V1V 1V7, Canada, ‡CIBIO, Centro de Investigação em Biodiversidade e Recursos Genéticos, Universidade do Porto, InBIO - Laboratório Associado, Campus Agrário de Vairão, 4485-661 Vairão, Portugal, §Departamento de Biologia, Faculdade de Ciências da, Universidade do Porto, Rua do Campo Alegre s/n, 4169-007 Porto, Portugal, ¶Fisheries, Wildlife and Conservation Biology Program, Department of Forestry and Environmental Resources, North Carolina State University, Raleigh, NC 27695, USA

## Abstract

With climate warming, the ranges of many boreal species are expected to shift northward and to fragment in southern peripheral ranges. To understand the conservation implications of losing southern populations, we examined range-wide genetic diversity of the snowshoe hare (*Lepus americanus*), an important prey species that drives boreal ecosystem dynamics. We analysed microsatellite (8 loci) and mitochondrial DNA sequence (cytochrome b and control region) variation in almost 1000 snowshoe hares. A hierarchical structure analysis of the microsatellite data suggests initial subdivision in two groups, Boreal and southwestern. The southwestern group further splits into Greater Pacific Northwest and U.S. Rockies. The genealogical information retrieved from mtDNA is congruent with the three highly differentiated and divergent groups of snowshoe hares. These groups can correspond with evolutionarily significant units that might have evolved in separate refugia south and east of the Pleistocene ice sheets. Genetic diversity was highest at mid-latitudes of the species' range, and genetic uniqueness was greatest in southern populations, consistent with substructuring inferred from both mtDNA and microsatellite analyses at finer levels of analysis. Surprisingly, snowshoe hares in the Greater Pacific Northwest mtDNA lineage were more closely related to black-tailed jackrabbits (*Lepus californicus*) than to other snowshoe hares, which may result from secondary introgression or shared ancestral polymorphism. Given the genetic distinctiveness of southern populations and minimal gene flow with their northern neighbours, fragmentation and loss of southern boreal habitats could mean loss of many unique alleles and reduced evolutionary potential.

**Keywords:** climate change, core-periphery, evolutionarily significant units, landscape genetics, *Lepus americanus*, phylogeography

Received 23 May 2013; revision received 5 May 2014; accepted 5 May 2014

## Introduction

Over the next century, North America's southern boreal forests are predicted to undergo rapid fragmentation and loss due to climate change and human activities

(IPCC 2007). Understanding the conservation implications of southern habitat loss for boreal species requires evaluating range-wide genetic structure of individual species and assessing the generality of these patterns across taxa. Specifically, insights into large-scale genetic diversity and population differentiation would clarify the relative importance of southern boreal populations as hotspots of diversity and evolutionarily significant

Correspondence: L. Scott Mills,  
E-mail: lsmills@ncsu.edu

units (ESUs; Moritz 1994), with implications for adaptive potential.

Distribution of genetic diversity and structure across a species' range reflects historical range contraction and recolonization from glacial refugia as well as current habitat fragmentation and dispersal. During the Quaternary ice ages (approximately 2.6 mya–present), the alternation of glacial and interglacial periods caused repeated changes in species' distributions. A frequently invoked 'southern refugia' model in phylogeography suggests that when ice sheets advanced, many boreal species persisted primarily in refugia in southern latitudes (Hewitt 1996). Interglacial periods enabled northward range expansion, with leading-edge populations carrying a subset of the genetic diversity of refugial populations. Simultaneously, range contraction to higher elevations in southern populations may have reduced connectivity and increased local diversification (Moritz *et al.* 2008). For some species in North America and Europe, a pattern of decreasing genetic diversity with increasing latitude ('southern richness, northern purity') may reflect a dominant influence of historical southern refugia on patterns of diversity (Pielou 1991; Hewitt 1996; Soltis *et al.* 1997).

The generality of the southern refugia model has been challenged for species with high dispersal and large contemporary ranges (Hewitt 2000; Provan & Bennett 2008). For many North American species, fossil evidence and phylogeographic studies have identified additional glacial refugia in eastern Beringia, the Canadian Arctic, coastal British Columbia, the Maritimes, and other northern locations (Soltis *et al.* 2006; Provan & Bennett 2008; Godbout *et al.* 2010). When expanding populations from separate refugia met in zones of secondary contact, they often created hotspots of genetic diversity (Provan & Bennett 2008). Geographic and genetic subdivision within major refugia further complicate genetic patterns (Gomez & Lunt 2007).

Contemporary gene flow also impacts genetic structure and diversity. Many boreal species have populations occupying peninsular habitat extensions into montane forests of the USA (Shugart *et al.* 2005). In addition to natural habitat fragmentation, these southern boreal forests are heavily impacted by logging and habitat conversion (Hansen *et al.* 2010; Powers *et al.* 2012), which could lead to differentiation in remnant habitats. The core-periphery hypothesis suggests that connected populations in the boreal range core should have higher genetic diversity than the fragmented populations of the southern periphery (Eckert *et al.* 2008). But while low gene flow and chronic genetic drift may reduce genetic diversity in peripheral populations, these processes may simultaneously facilitate genetic differentiation and preserve unique alleles (Eckert *et al.* 2008).

Given the complex interplay of forces that shape intraspecific distribution of genetic diversity, what are the consequences of losing southern boreal populations? In this study we examined range-wide genetic diversity of the snowshoe hare (*Lepus americanus*) to address this question.

Snowshoe hares are important prey for most boreal carnivores, structuring food web dynamics as strong interactors (Krebs *et al.* 2001). Fossil evidence suggests the persistence of snowshoe hares in extensive refugia south of the ice sheets and in the northern refugium of Beringia during the Last Glacial Maximum (LGM; FAUNMAP Working Group 1994). Snowshoe hare populations near historical Beringia and in Montana harbour high genetic diversity and are genetically differentiated from each other (Burton *et al.* 2002). Morphological differences among snowshoe hare populations in and around the Pacific Northwest suggest genetically differentiated populations (Dalquest 1942).

We analysed mtDNA and microsatellite data throughout the contemporary range of snowshoe hares to test the hypotheses that: (i) extant snowshoe hare populations derive from Beringia and southern refugia; and (ii) snowshoe hare populations near the core of the range exhibit higher genetic diversity than populations near the periphery. We predict highest genetic diversity and uniqueness in the species' southern range and near the Alaska-Yukon border, that is, in likely refugia, with reduced diversity in range-edge populations outside of these areas.

We then discuss how anticipated loss of southern boreal habitats might affect snowshoe hare genetic diversity. First, we determine whether multiple snowshoe hare ESUs (*sensu* Moritz 1994) are warranted. Second, we examine genetic diversity and uniqueness across a latitudinal gradient, with particular focus on populations below 49°N, the approximate southernmost extent of the LGM. Many scenarios of climate change predict the climate envelope for North America's boreal ecosystem will shift north of 49°N within a century (Koven 2013). Finally, we discuss similarities in findings between snowshoe hares and other North American hare species.

## Materials and methods

We analysed 975 snowshoe hare samples from 16 U.S. states and 12 Canadian provinces and territories (Appendix S1, Supporting information). Nearly all samples were ear tissue collected from road kill, game harvests, and live-trapping during 1989–2010. Ten samples were faecal pellets collected in Isle Royale, Michigan, in 2009. Eleven samples were tissue from specimens collected near Vancouver, British Columbia, from 1929 to 1970, held at the University of British Columbia Cowan

Museum. For phylogenetic analyses, we additionally analysed one white-tailed jackrabbit (*Lepus townsendii*) tissue sample obtained from GenBank (Accession no. AY292729; Matthee *et al.* 2004) and seven black-tailed jackrabbit (*L. californicus*) tissue samples collected from three U.S. states (California, New Mexico, Nevada).

### Microsatellite analysis

Samples were grouped into populations on the basis of two geographic criteria: (i) no potential genetic barriers such as large lakes or rivers, mountain ranges, or non-forested regions bisecting populations (Burton *et al.* 2002; Shafer *et al.* 2010b); and (ii) a maximum of 260 km between any two samples in a population. The second criterion represents a coarse spatial scale much greater than the distance hares disperse (up to ~20 km, Gillis & Krebs 1999) but within the scale of reported gene flow in northern snowshoe hare populations (~600 km, Burton *et al.* 2002). After grouping samples, we limited genetic analyses to groups with at least seven samples. This minimum threshold was arbitrary, but has precedence in other population genetic studies (Schwartz *et al.* 2003; Tracy & Jamieson 2011).

We selected eight polymorphic microsatellite markers developed in the European rabbit, *Oryctolagus cuniculus*, and successfully used with snowshoe hares (Burton *et al.* 2002; Schwartz *et al.* 2007): 7L1D3 (Korstanje *et al.* 2003); SAT02, SAT12, SAT13, SAT16 (Mougel *et al.* 1997); SOL08, SOL30 (with 'GTGTCTT' tail added) (Rico *et al.* 1994); and SOL33 (SurrIDGE *et al.* 1997) (Appendix S2, Supporting information). DNA extraction and genotyping methods are detailed in Appendix S3 (Supporting information).

Allelic dropout and false allele rates were calculated with 10 000 search iterations in Pedant version 1.0. (Johnson & Haydon 2007). For each population, we used Genepop version 4.0.11 (Rousset 2008) to test for Hardy-Weinberg Equilibrium (HWE) and linkage disequilibrium. Markov chain parameters for exact tests were set at 10 000 dememorizations, 100 batches, and 5000 iterations per batch (Raymond & Rousset 1995). We used the false discovery rate approach (FDR; Benjamini & Hochberg 1995) in the R software package 'fdrtool' (Strimmer 2008; <http://cran.r-project.org/>) to correct for multiple significance testing type I error. Potential null alleles and scoring errors due to stuttering and allelic drop-out were identified by Monte Carlo simulation in Micro-Checker version .2.2.3 (van Oosterhout *et al.* 2004).

We performed Bayesian analyses in STRUCTURE version 2.3.3 (Pritchard *et al.* 2000) and Geneland version 4.0.3 (Guillot *et al.* 2005) to partition microsatellite data into genetic clusters, and to assign individuals to their likely cluster of origin. In STRUCTURE, we

applied an admixture model with the 'locprior' option, using a burn-in period of 20 000 generations and 100 000 MCMC iterations after burn-in. We compared results from correlated vs. uncorrelated allele frequency models. To check for MCMC consistency, we performed 20 replicates for each *K* (number of clusters) from 1 to 40. The most likely *K* value was determined in Structure Harvester version 0.6.93 (Earl & vonHoldt 2011) as the likelihood model with the highest  $\Delta K$  (Evanno *et al.* 2005), unless the maximum  $\ln P(D)$  was for *K* = 1 (which would indicate no substructure). In the presence of substructure, the  $\Delta K$  method detects the highest hierarchical structure (Evanno *et al.* 2005). We assigned each individual to its most probable cluster and repeated the analysis for each cluster separately, until further substructure could not be detected. Cluster assignment was based on outcomes from the run with highest  $\ln P(D)$  among 20 replicates. Following Coulon *et al.* (2008), only individuals with at least 60% membership in a cluster were included in subsequent analyses. For each subsequent analysis, model parameters remained the same, but maximum *K* was set at one greater than the number of sampled populations in that cluster.

In Geneland, we evaluated results from three spatially explicit model combinations. We examined both correlated and uncorrelated allele frequency models without filtering null alleles. We additionally examined an uncorrelated frequency model while filtering null alleles. For each of the three model combinations we ran 20 independent replicates of 1 000 000 MCMC iterations with a thinning of 1000 and burn-in of 200 000 iterations. *K* was allowed to vary from 1 to 40. Following program recommendations for our sample size, we set maximum rate of Poisson process = 853 and maximum number of nuclei in the Poisson-Voronoi tessellation = 2559. We allowed a 15-km uncertainty in spatial coordinates. MCMC convergence was assessed for each model combination by comparing estimated *K* and cluster assignments across replicate runs.

GENALEX version 6.3 (Peakall & Smouse 2006) was used to calculate number of alleles and expected heterozygosity (Nei 1978) for each population. For all pairs of populations, we estimated Nei's *D* (Nei 1972) and Weir & Cockerham's (1984)  $F_{ST}$ , with the latter calculated in ARLEQUIN version 3.5.1.2 (Excoffier *et al.* 2005). Significance was determined with 1000 permutations of samples among populations and FDR correction for multiple comparisons.

We used rarefaction, implemented in HP-RARE version 1.0 (Kalinowski 2005), to calculate private allelic richness (PAR) for each population, standardized to the smallest sample size (seven individuals) in this study. To minimize biases due to uneven sampling

density, all populations within a 350 km radius were excluded from the calculation of PAR for each sampled population (we also tested 500 km and results were similar; data not shown). We examined scatterplots of genetic metrics against latitude and longitude to identify signatures of genetic drift at the current range periphery and to understand geographic patterns of diversity.

### *Mitochondrial DNA analysis*

We amplified a 468 bp fragment of the mitochondrial control region (CR) with primers LCRSEQ (Melo-Ferreira *et al.* 2007) and LepD2H (Pierpaoli *et al.* 1999) in all snowshoe hare samples. A fragment with 633 bp of the cytochrome b (Cytb) gene was also sequenced in a subset of 80 snowshoe hare and seven black-tailed jackrabbit samples, using primers LGCYF (Alves *et al.* 2003) and LCYTBR (Melo-Ferreira *et al.* 2005), as detailed in Appendix S3 (Supporting information). The Cytb subset comprised at least one snowshoe hare sample from each population and additional samples from regions of high CR genetic structure. We visually aligned sequences in CodonCode Aligner version 3.5.4 (CodonCode Corporation, Dedham, MA, USA).

Phylogenetic trees were constructed in BEAST version 1.7.4 (Drummond *et al.* 2012) based on the Cytb gene, which has a slower mutation rate and thus lower tendency than CR for homoplasy over long timescales (Baker & Marshall 1997). We used jModelTest version 2.1.3 (Darriba *et al.* 2012) and the Bayesian information criterion to assess the best-fit model of sequence evolution. Posterior probabilities were determined from three independent runs of 250 million generations, using the selected mutation model, the Yule tree prior and a random local clock (Drummond & Suchard 2010), excluding the initial 10% of each run as burn-in. The stability of the runs and convergence of the MCMC were assessed with Tracer version 1.5 (<http://beast.bio.ed.ac.uk/Tracer>). Results from the three runs were concatenated in LogCombiner version 1.7.4 and trees annotated using TreeAnnotator version 1.7.4. The annotated phylogenetic tree and posterior probability estimates were visualized in FigTree version 1.4.0 (<http://tree.bio.ed.ac.uk/software/figtree/>). To estimate lineage divergence times, we used a mutation rate of 0.02 substitutions per site per million years (Brown *et al.* 1979), which has been used to estimate divergence in other hare studies (Pierpaoli *et al.* 1999; Melo-Ferreira *et al.* 2007).

The demographic history of the major mtDNA lineages was inferred from control region sequences using the Bayesian Skyline Plot (BSP) (Drummond *et al.* 2005) implemented in BEAST. Three replicate runs of 100

million generations were performed using the appropriate mutation models (for Boreal, TrN+I+G; for Greater Pacific Northwest and U.S. Rockies, HKY+G) selected using the procedure described above and a random local clock (Drummond & Suchard 2010). Tracer version 1.5 was used to assess stability of the MCMC and the initial 10% of each run was discarded as burn-in. We used LogCombiner version 1.7.4 to concatenate results of the three replicate runs. A CR mutation rate of 0.156 substitutions per site per million years (derived from Melo-Ferreira *et al.* 2007) was used to calibrate the BSP.

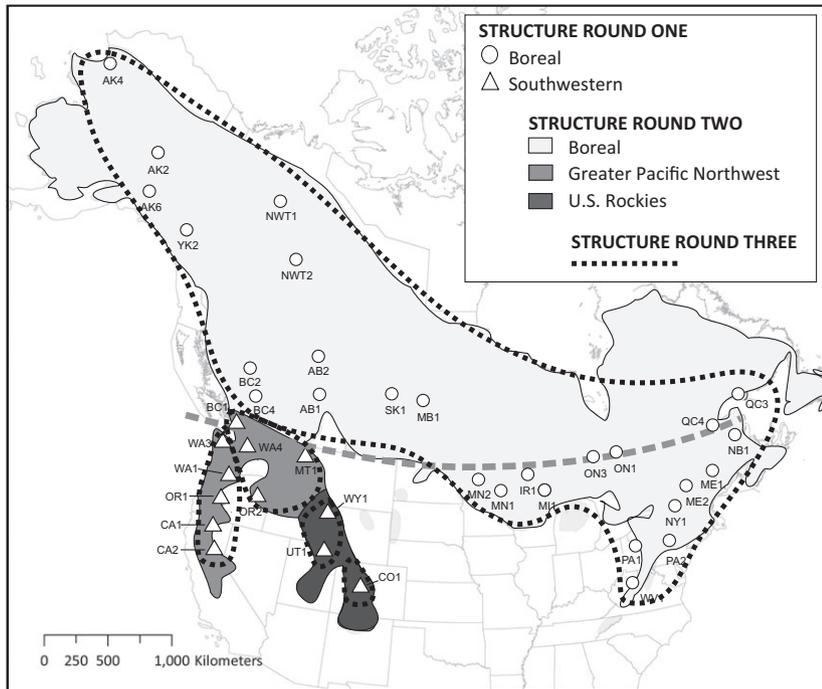
We examined within-lineage structure with the CR gene, because its fast rate of evolution makes it suitable for intraspecific studies (Vigilant *et al.* 1991). An unrooted median-joining network (NETWORK version 4.5.1.6, <http://www.fluxus-engineering.com/>) was generated from CR haplotypes identified in DnaSP version 5.10 (Librado & Rozas 2009). Transversions were weighted three times as high as transitions, following software recommendations. For  $K = 1-10$ , SAMOVA version 1.0 (Dupanloup *et al.* 2002) identified the partitioning of CR haplotype variance due to differences among groups. We ran SAMOVA with 500 initial population partitions and 10 000 iterations for each  $K$ . Significance of variance components was evaluated by 1000 permutations of populations among groups.

We used ARLEQUIN v.3.5.1.2 (Excoffier *et al.* 2005) to calculate haplotype and nucleotide diversities. As with microsatellite data, scatterplots were used to assess latitudinal and longitudinal patterns in genetic diversity. To evaluate genetic differentiation, we calculated pairwise control region  $F_{ST}$ . ARLEQUIN v.3.5.1.2 was used to determine significance of tests with 10 000 bootstraps and FDR control for multiple comparisons.

## Results

### *Microsatellite analysis*

With an average of 2.2 PCR replicates per sample, we successfully genotyped eight microsatellite loci for 922 snowshoe hares. The mean allelic dropout rate per allele was 0.0070 and mean false allele rate was 0.0035, for all loci combined. After excluding populations with <7 individuals, 853 samples in 39 populations remained for analyses (Fig. 1). Only 4% of 312 population-loci combinations significantly deviated from Hardy-Weinberg Equilibrium, generally due to heterozygote deficit. Slightly over 5% of 1026 tests for linkage disequilibrium were significant. Micro-checker identified potential null alleles in 8% of 312 population-loci tests. However, null alleles and deviations from HWE were not associated more frequently with any particular locus, and genotypic disequilibrium was not consistently attributed



**Fig. 1** Sampling locations and geographic distribution of major snowshoe hare microsatellite clusters, as defined by STRUCTURE version 2.3.3 hierarchical analysis (Pritchard *et al.* 2000). The first run of STRUCTURE distinguished Boreal (circles) from southwestern (triangles) populations. The second run further split the southwestern cluster (grey shades). By the third run, five distinct genetic clusters were identified (dotted ovals). The approximate southernmost latitude of the LGM is marked with a grey dashed line. Study results suggest extant snowshoe hare populations likely expanded from refugia south of this latitude and from the current eastern hare range.

to a particular locus pair. Therefore we retained all loci for subsequent analyses.

STRUCTURE analyses identified hierarchical population division (Fig. 1 and Appendix S4, Supporting information). In the first round of STRUCTURE runs, the highest likelihood model ( $K = 2$ ) identified a Boreal cluster comprising the entire northern and eastern range of the species, and a southwestern cluster comprising remaining populations. The second round of STRUCTURE further splits the Boreal cluster into two subclusters. However, proportion membership in the subclusters transitioned from west to east (Appendix S5, Supporting information), suggesting an effect of isolation by distance rather than historical isolation (Meirmans 2012). A Mantel test (Mantel 1967), following Rousset's (1997) method, confirmed a significant correlation between geographic and genetic distance in the Boreal cluster ( $P < 0.001$ ). Because other analyses and markers also supported a single Boreal cluster, we did not continue STRUCTURE analyses to further subset the Boreal cluster. In contrast to the Boreal cluster, the second round of STRUCTURE clearly divided the southwestern group into two genetic clusters, corresponding to the Greater Pacific Northwest region and to the U.S. Rockies. Further rounds of the hierarchical STRUCTURE analysis subdivided the Greater Pacific Northwest and U.S. Rockies groups into many subclusters. Ultimately, by the fifth round of analysis, all southwestern populations were identified as distinct subclusters except for WA1 and WA4 in Washington. For these two populations, some individuals could not

be assigned to a cluster with at least 60% probability, and other individuals grouped with other populations. Hierarchical cluster patterns were identical for the correlated and uncorrelated allele frequency models.

Using Geneland, all replicates of the uncorrelated frequency models (with and without filtering null alleles) consistently identified a single Boreal cluster and 4–7 distinct clusters in the species' southwestern range. With the uncorrelated model and null alleles filtered, the highest mean posterior density across 20 replicates was obtained for  $K = 5$ , with clusters almost identical to those identified from the first three rounds of STRUCTURE hierarchical analysis (Appendix S6, Supporting information). All replicates of the correlated frequency model in Geneland inferred 39–40 genetic clusters, likely due to known instabilities of this model in the presence of isolation by distance (Guillot 2008).

Measures of  $F_{ST}$  and Nei's  $D$  were highly correlated across populations ( $r = 0.93$ ,  $P < 0.001$ ). We found high  $F_{ST}$  pairwise estimates ( $>0.20$ ) between the three genetic clusters identified in the first two rounds of STRUCTURE. These clusters were the most congruent across markers (microsatellite and mtDNA) and analyses. Pairwise  $F_{ST}$  was high within the Greater Pacific Northwest and U.S. Rockies clusters, but was usually below 0.20 within the Boreal cluster (Appendix S7, Supporting information).

Most snowshoe hare populations were characterized by high genetic diversity (Table 1). On average, populations in the Boreal cluster exhibited the highest allelic richness and heterozygosity, but the lowest uniqueness

**Table 1** Microsatellite diversities averaged across 8 loci for each of 39 sampled populations. Populations are grouped into three genetic clusters identified by the first two rounds of STRUCTURE hierarchical analysis

Population	N	A	AR	$H_O$	$H_E$	PAR
AB1	9	7.00	6.20	0.69	0.76	0.13
AB2	18	8.63	5.95	0.69	0.77	0.14
AK2	28	7.50	4.79	0.56	0.62	0.02
AK4	15	5.75	4.63	0.56	0.64	0.07
AK6	9	4.75	4.38	0.61	0.64	0.19
BC2	25	7.88	5.09	0.57	0.67	0.07
BC4	9	4.75	4.33	0.56	0.65	0.08
IR1	10	5.88	5.13	0.66	0.71	0.05
MB1	13	6.50	5.12	0.59	0.71	0.10
ME1	40	9.75	5.89	0.72	0.77	0.06
MI1	8	6.50	6.10	0.72	0.80	0.33
MN1	34	9.63	5.62	0.58	0.73	0.07
MN2	12	7.50	5.97	0.70	0.79	0.04
NB1	20	9.00	5.86	0.68	0.74	0.17
NWT1	9	6.50	5.83	0.69	0.74	0.11
NWT2	18	7.63	5.59	0.67	0.72	0.01
NY1	13	6.75	5.58	0.70	0.79	0.10
ON1	11	7.00	5.86	0.67	0.75	0.02
ON3	19	9.25	5.85	0.66	0.75	0.15
PA1	10	5.25	4.63	0.63	0.65	0.15
PA2	13	6.63	5.23	0.70	0.68	0.07
QC3	20	8.50	5.80	0.70	0.77	0.09
QC4	17	7.00	5.38	0.71	0.76	0.05
SK1	8	6.13	5.78	0.56	0.72	0.20
VT1	10	5.63	4.97	0.70	0.73	0.03
WV1	14	4.50	3.90	0.50	0.63	0.10
YK2	30	8.50	5.10	0.62	0.65	0.11
<i>Boreal</i>	<i>Total</i>	<i>7.05 (1.50)</i>	<i>5.35 (0.61)</i>	<i>0.64 (0.06)</i>	<i>0.72 (0.06)</i>	<i>0.10 (0.07)</i>
	<i>N = 442</i>					
BC1	15	4.25	3.30	0.46	0.51	0.02
CA1	12	4.88	4.16	0.59	0.61	0.13
CA2	7	3.63	3.63	0.48	0.52	0.01
MT1	100	12.13	5.58	0.70	0.73	0.17
OR1	32	9.00	5.29	0.64	0.68	0.23
OR2	17	5.88	4.48	0.62	0.61	0.23
WA1	30	9.25	5.91	0.71	0.77	0.38
WA3	9	5.13	4.64	0.71	0.65	0.29
WA4	29	9.38	5.73	0.67	0.76	0.35
<i>Greater Pacific Northwest</i>	<i>Total</i>	<i>7.06 (2.94)</i>	<i>4.75 (0.94)</i>	<i>0.62 (0.09)</i>	<i>0.65 (0.10)</i>	<i>0.20 (0.13)</i>
	<i>N = 251</i>					
CO1	58	7.75	4.25	0.55	0.57	0.14
UT1	25	4.88	3.66	0.50	0.54	0.27
WY1	77	7.25	4.06	0.53	0.55	0.10
<i>U.S. Rockies</i>	<i>Total</i>	<i>6.63 (1.53)</i>	<i>3.99 (0.30)</i>	<i>0.53 (0.03)</i>	<i>0.55 (0.02)</i>	<i>0.17 (0.09)</i>
	<i>N = 160</i>					

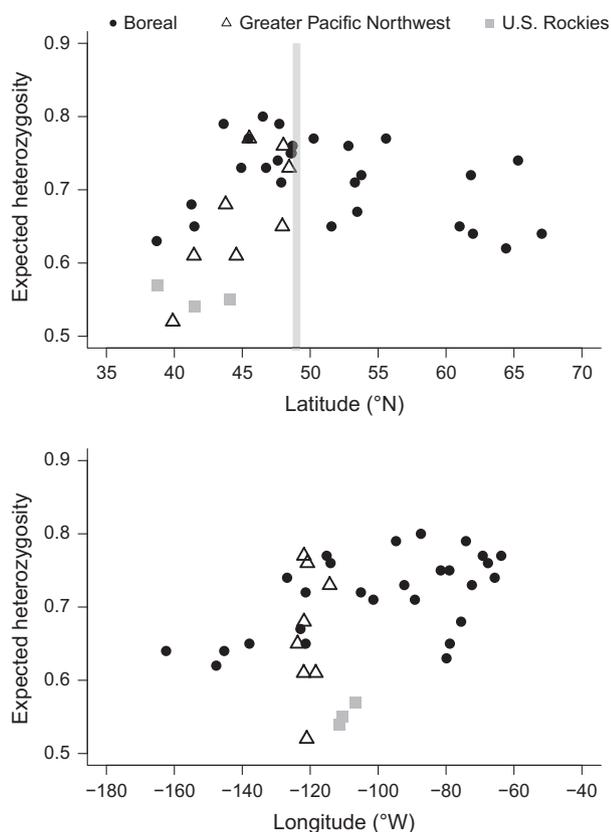
N, number of individuals; A, number of different alleles; AR, allelic richness;  $H_O$ , observed heterozygosity;  $H_E$ , expected heterozygosity; PAR, population private allelic richness.

Cluster averages and standard deviations (in parentheses) are italicized.

(PAR). Greater Pacific Northwest populations exhibited high diversity and the highest uniqueness of the three major clusters.

Genetic diversity was highest at mid-latitudes (i.e. near 49°N latitude; Fig. 2 and Appendix S8, Supporting

information) and increased from west to east across the species' range. For the Greater Pacific Northwest cluster, PAR increased with latitude up to 49°N latitude (Fig. 3). There were no apparent longitudinal trends in PAR.

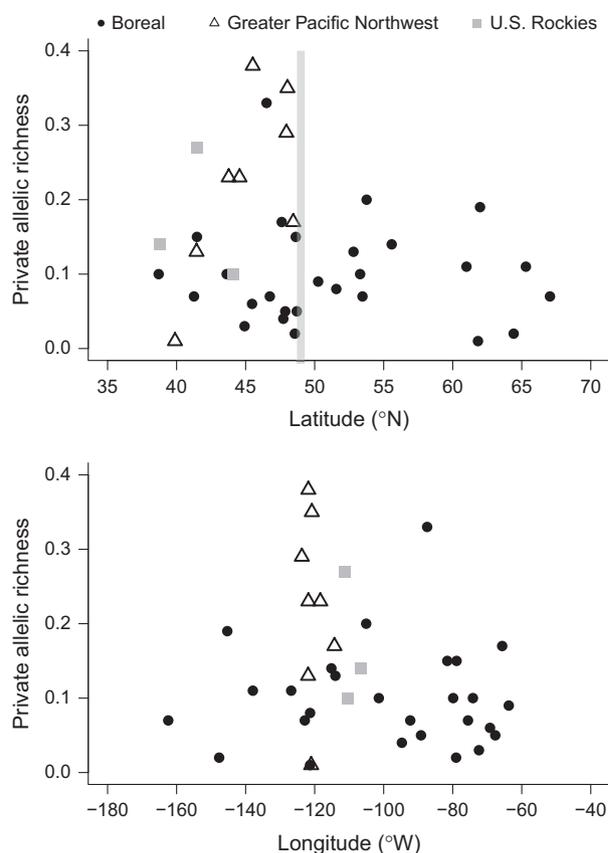


**Fig. 2** For each sampled population, expected heterozygosity plotted against latitude (top) and longitude (bottom). The grey vertical bar marks the approximate southernmost latitude of the LGM.

#### Mitochondrial DNA analysis

The final data set for CR analyses comprised 893 snowshoe hare samples represented by 365 haplotypes. For phylogenetic tree construction, the subset of 80 snowshoe hare Cytb sequences comprised 43 haplotypes.

The best-fit model of nucleotide substitution for Cytb phylogeny was HKY+G. Three highly divergent lineages were identified (Fig. 4), broadly corresponding with the major genetic clusters identified in the first two rounds of STRUCTURE analysis of microsatellites (Fig. 1 and Appendix S4, Supporting information). MtDNA analysis also identified two sublineages (with >95% posterior probability) that corresponded with the finer scale splitting of southwestern populations from hierarchical STRUCTURE analysis: (i) WA3 in Olympic National Park, Washington, was a sublineage of the Greater Pacific Northwest lineage; and (ii) CO1 in Gunnison, Colorado, was a sublineage of the U.S. Rockies lineage. Within the Boreal lineage, a basal group comprised samples from populations near the lineage's southern range (MT1, Montana; MB1, Manitoba; ON1, Ontario; Fig. 4). The Cytb topology indicated that

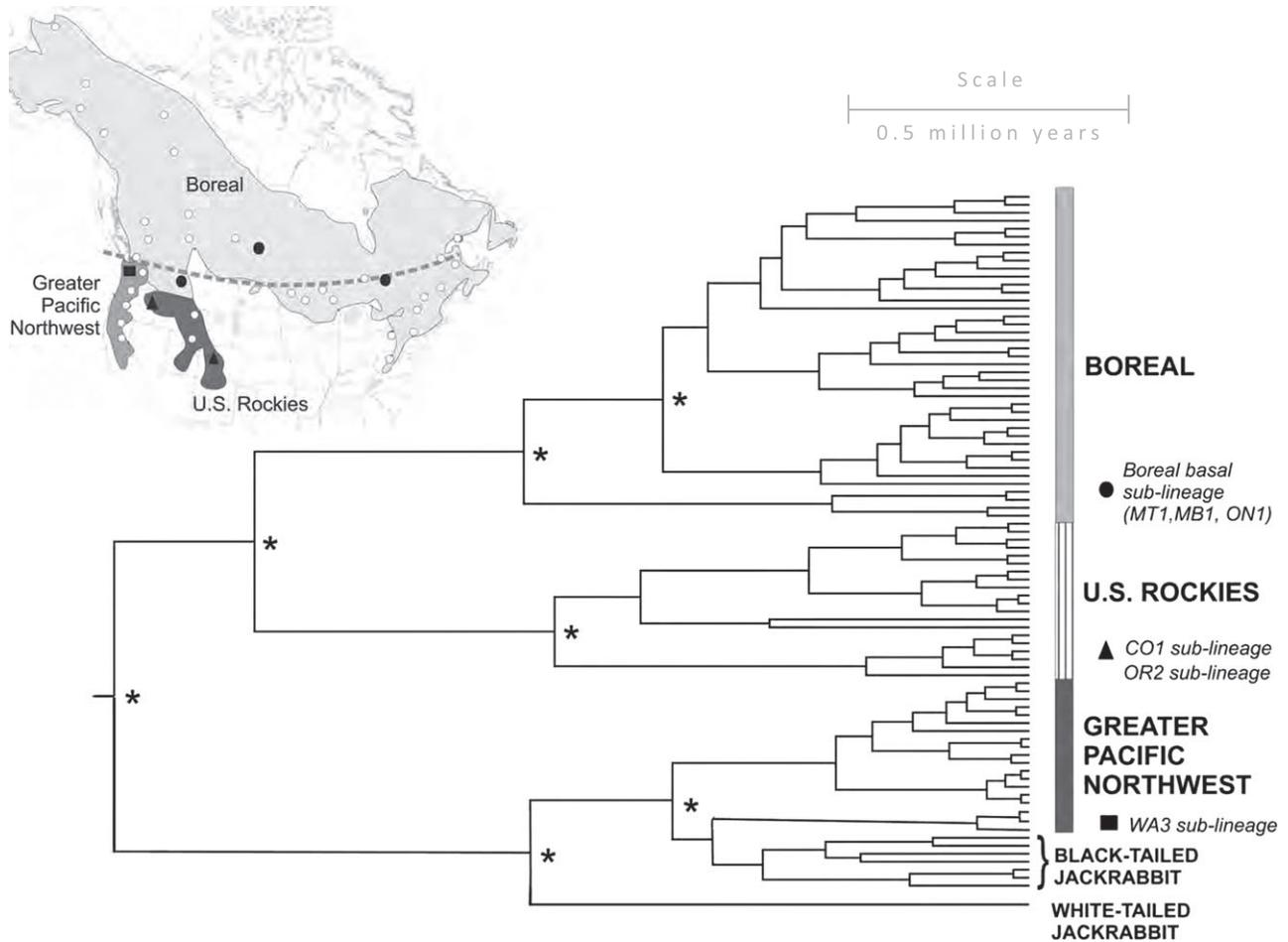


**Fig. 3** For each sampled population, private allelic richness (PAR) plotted against latitude (top) and longitude (bottom). The grey vertical bar marks the approximate southernmost latitude of the LGM.

snowshoe hares in the Greater Pacific Northwest lineage are more closely related to black-tailed jackrabbits than to other snowshoe hare populations.

Divergence estimation between the Boreal and U.S. Rockies snowshoe hare lineages is 1.30 mya (95% CI 0.88–1.74 mya). The CO1 sublineage split off from the U.S. Rockies major lineage more recently (0.78 mya; 95% CI 0.42–1.18 mya). The clade comprising most of the snowshoe hares in the Greater Pacific Northwest lineage diverged from BTJR about 0.59 mya (95% CI 0.34–0.90 mya; Fig. 4). The Bayesian skyline plot provided strong evidence of a recent demographic expansion of the Boreal lineage (Fig. 5), whereas expansion of the Greater Pacific Northwest and U.S. Rockies groups was not supported.

NETWORK and SAMOVA analyses, based on the CR gene, accorded with the Cytb phylogeny. In an unrooted median-joining network, the Cytb lineages and sublineages were reciprocally monophyletic and separated from each other by  $\geq 10$  CR base pair substitutions (Appendix S9, Supporting information). By these criteria, the OR2 population of Malheur National Forest,



**Fig. 4** Snowshoe hare phylogenetic relationships, as constructed in BEAST version 1.7.4 (Drummond *et al.* 2012), based on the *Cytb* gene. Asterisks (\*) indicate lineages and sublineage divisions with  $\geq 95\%$  posterior probability support. Map shows the geographic distribution of the three major *Cytb* lineages. Filled symbols identify sublineages.

Oregon, was also identified as a distinct sublineage of the U.S. Rockies lineage.

Differences between the Greater Pacific Northwest lineage and all other snowshoe hare lineages explained 77% of total genetic variation in the *CR* gene (SAM-OVA; Dupanloup *et al.* 2002). If the Greater Pacific Northwest snowshoe hare lineage is introgressed from black-tailed jackrabbits, this deep genetic division when  $K = 2$  is an artefact of interspecific hybridization rather than snowshoe hare demographic history. Therefore, we also analysed the data with the Greater Pacific Northwest lineage excluded. In this analysis, 66% of variation was explained by differences between four groups ( $K = 4$ ,  $P < 0.001$ ): the Boreal lineage, U.S. Rockies lineage, and the sublineages CO1 and OR2. Control region pairwise  $F_{ST}$  was high between and within all lineages except within the Boreal lineage (Appendix S10, Supporting information).

Only two populations (MT1 and WA4; Fig. 1) contained haplotypes that could be ascribed to more than

one lineage, with both showing ties to Boreal and Greater Pacific Northwest lineages. Most populations were characterized by high haplotype and nucleotide diversities, with exceptions in British Columbia (BC1, near Vancouver) and in southern populations (CA1 and CA2 in California; UT1 in Utah; WV1 in West Virginia; Appendix S11, Supporting information). Haplotype and nucleotide diversities increased with latitude up to  $\sim 49^\circ\text{N}$  latitude (Appendices S12 and S13, Supporting information). MtDNA diversity did not exhibit any clear longitudinal pattern.

## Discussion

In this range-wide study of a species distributed across boreal North America, we found that snowshoe hares formed three major genetic groups with well-defined distributions, coincident with patterns observed in other North American boreal species (Arbogast 1999; van Els *et al.* 2012). The entire northern and eastern range of the

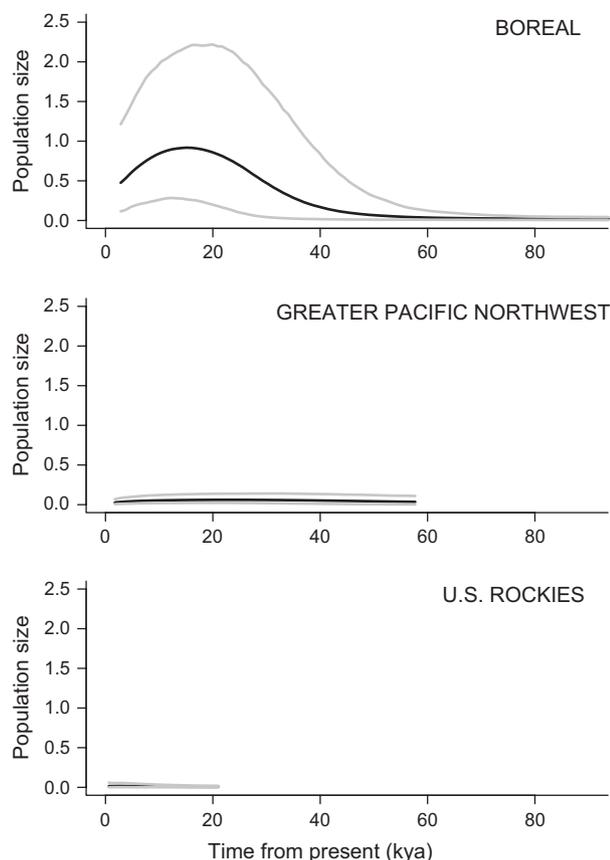


Fig. 5 Bayesian skyline plots (BEAST version 1.7.4; Drummond *et al.* 2012) for three snowshoe hare genetic lineages. Relative population sizes are in units of  $N_e \times$  mutation rate. Grey lines represent the 95% CI.

snowshoe hare, spanning 6000 km across Canada and the eastern U.S., constituted a single Boreal group characterized by high genetic diversity and gene flow. Two geographically confined groups—in the Greater Pacific Northwest and U.S. Rockies—exhibited lower gene flow and high genetic uniqueness. The three major groups are coherent from microsatellite and mtDNA analyses. Both markers further identified genetic subdivision within the Greater Pacific Northwest and U.S. Rockies, of which the separation of CO1 (Colorado) from the U.S. Rockies group was congruent across all markers and analyses. Modern populations of snowshoe hares likely derived from refugial populations that persisted through the Quaternary ice ages in eastern and southern refugia.

We found high genetic diversity in most sampled populations, but reduced diversity at current range edges, especially for populations at the species' fragmented southern edge. Southern range populations below 49°N had high genetic uniqueness with minimal gene flow with their northern neighbours, suggesting

snowshoe hares could lose considerable genetic diversity if southern boreal habitats are lost.

#### *Evolutionary history and refugial origin*

This work revealed strong genetic structure at different hierarchical levels and a remarkable coincidence of the inference of three major geographically explicit groups of snowshoe hares based on mtDNA sequences and microsatellite data. Further, these markers also coincide in the suggestion of additional genetic fragmentation in the species' southwestern range. Based on the mtDNA phylogeny, we estimated divergence of the three major groups to be 1.30–0.78 mya, long before the height of the LGM ~18 kya. Regional mixing among groups was sufficiently low during subsequent interglacial warm periods, including the current one, that deep genetic divisions are still maintained in the mtDNA and microsatellite data.

Many co-occurring forest species for which continent-wide genetic data are available share this phylogeographic pattern—a large genetic cluster across Canada and the eastern U.S. and one or more smaller genetic clusters in the western USA. Examples include the gray jay (*Perisoreus canadensis*, van Els *et al.* 2012), northern flying squirrel (*Glaucomys spp.*, Arbogast 1999), black bear (*Ursus americanus*, Wooding & Ward 1997) and hairy woodpecker (*Picoides villosus*, Klicka *et al.* 2011). Genetic groups in these species diverged an estimated 2.97–0.69 mya, a range that encompasses our divergence estimates for the snowshoe hare.

*Boreal snowshoe hare lineage.* The height of the LGM in North America occurred ~18 kya, and by 6 kya, the glaciers had largely disappeared (Pielou 1991). Given the Boreal lineage diverged from other snowshoe hare lineages an estimated 1.30 mya, colonization of newly available boreal habitats after the LGM must have occurred primarily from refugial populations within the Boreal lineage. We had hypothesized a Beringian refugium for snowshoe hares, as reported for several other North American boreal species (Shafer *et al.* 2010b), but we did not find genetic diversity or uniqueness patterns indicative of a major Beringian refugium for snowshoe hares. A few snowshoe hare fossils are documented from Alaska and Yukon from 20 to 10 kya, but the majority of hare fossils from this period are from the lower 48 U.S. states (FAUNMAP Working Group 1994). Thus, any relict snowshoe hare populations that survived the LGM in Beringia may have been too small or isolated to be heavily represented in contemporary snowshoe hare genetic structure.

Instead, genetic patterns suggest that the Boreal lineage primarily expanded from refugia near the southern

edge of the ice sheets and from eastern refugia. This idea is supported by the basal position of snowshoe hare *Cytb* haplotypes sampled from locations close to current southern limits of the Boreal lineage. In addition, microsatellite diversity was highest in eastern populations of the Boreal lineage. Results are consistent with fossil pollen data, which indicate that at the LGM, boreal forests in North America persisted in at least two major pockets—the Pacific Northwest and the southeastern USA (Williams *et al.* 1993).

The overall high genetic diversity through much of the Boreal lineage, and the significant pattern of IBD across the Boreal lineage range, suggests cross-continental expansion may have proceeded slowly or from a broad refugial front (Hewitt 1996). Signals of demographic expansion revealed in the Bayesian skyline plot indicate Boreal lineage expansion may have begun ~48 000 years ago.

*Greater Pacific Northwest and U.S. Rockies snowshoe hare lineages.* The Greater Pacific Northwest and U.S. Rockies lineages occur in the species' southwest range, which was largely ice free during the Pleistocene. The high genetic uniqueness and strong genetic subdivisions in mtDNA of these lineages indicate that they arose from at least two discrete refugia. Comparative phylogeographic studies have identified the northwestern USA as an area of exceptionally high genetic differentiation for boreal and temperate species, due to the complex physiography of the region and its relative stability as a glacial refugium (Soltis *et al.* 1997; Swenson & Howard 2005; Shafer *et al.* 2010b). Our Bayesian estimates of temporal fluctuations of effective population size suggested these evolutionary groups remained relatively stable through evolutionary time (Fig. 5).

The mtDNA analyses indicate the Greater Pacific Northwest snowshoe hares are more closely related to black-tailed jackrabbits than to other snowshoe hares. Two competing hypotheses may explain this result: (i) mitochondrial DNA introgression (through hybridization) from *Lepus californicus* into *Lepus americanus* in the southwestern range of the latter or (ii) retention of an ancestral polymorphism shared between the two species (Moore 1995). Extensive mtDNA introgression occurs among other species of hares, resulting from ancient or current contacts among species and sometimes causing extensive replacements of lineages (Alves *et al.* 2008). Even though the geographic restriction of the shared variants and the remarkably close phylogenetic relationship with current *L. californicus* variants support the introgression hypothesis, the inference of such phenomena would require reconstruction of the speciation history of the taxa, using genealogical information from nuclear loci (Melo-Ferreira *et al.* 2012). It is

nevertheless important to note that if introgression caused this interspecific sharing of lineages, it was remarkably pervasive and may have completely replaced the mtDNA variation in the Greater Pacific Northwest evolutionary group identified using microsatellites.

#### *Genetic diversity of core vs. peripheral populations*

Genetic diversity of snowshoe hares was highest in mid-latitude populations, near the southernmost edge of the LGM. From here, diversity clearly decreased towards the south and less dramatically towards the north. The southern range edge for snowshoe hares is highly impacted by natural and anthropogenic habitat fragmentation (Hansen *et al.* 2010). The observed genetic pattern is consistent with the core-periphery hypothesis, with populations in the fragmented southern periphery exhibiting the lowest genetic diversity and gene flow. Further, high amplitude population fluctuations may promote gene flow and genetic diversity (Ehrich *et al.* 2009). Snowshoe hare populations across their northern range undergo large population cycles, whereas southern populations may have reduced cyclicity (Hodges 2000).

#### *Anticipated genetic consequences of southern population loss*

Our study provides important insights on how potential loss of southern hare populations (below 49°N) may affect genetic diversity. The strong genetic subdivisions and uniqueness of snowshoe hare populations suggest that anticipated fragmentation and loss of these habitats due to climate change and human activities may greatly reduce overall species genetic diversity, with possible negative implications for future adaptive potential. For example, we identified at least three snowshoe hare evolutionarily significant units (ESUs), using Moritz's (1994) criteria of reciprocal monophyly for mtDNA and significant divergence in the frequencies of nuclear alleles. Two ESUs occurred wholly in the species' southern range. Three snowshoe hare sublineages, reciprocally monophyletic and separated from each other by  $\geq 10$  CR base pair substitutions, were also found in the southern range. Additional ESUs may occur in parts of the southern range not sampled in this study: for example, we did not sample hares in New Mexico or in northern Idaho, an area with high endemism hypothesized to be the 'Clearwater refugium' (Daubenmire 1975; Soltis *et al.* 1997).

A limitation of this study is its reliance on neutral genetic variation, without complementary information on adaptive potential, for identifying ESUs (Funk *et al.*

2012). We identified ESUs on the basis of Moritz's definition because it can be operationally applied from neutral genetic markers (de Guia & Saitoh 2007). Other definitions of ESU emphasize conserving adaptive variation, by incorporating adaptive genetic variation, life history traits, morphology and species distribution (Ryder 1986; Vogler & DeSalle 1994). An additional question that should be addressed is, 'How much would loss of southern populations impact the species' ability to adapt to global warming?' Such studies would require evaluation of quantitative genetic trait variations directly linked to traits with adaptive value under altered climate regimes. For snowshoe hares, adaptive variation may include phenology of seasonal coat colour moult confronting decreased snow pack, especially in the southern part of the range (Mills *et al.* 2013).

Concomitant with the predominantly southern distribution of ESUs and sublineages, a large proportion of snowshoe hares' neutral private allelic richness (PAR) occurs in the U.S. Rockies and Greater Pacific Northwest, where isolation and relative stability over evolutionary time were likely responsible for their accumulation of mutations and unique genetic structure. On average, populations in the U.S. Rockies and Greater Pacific Northwest lineages had almost twice the PAR of populations in the Boreal lineage. In contrast to the highly connected Boreal populations, loss of a population in the U.S. Rockies and Greater Pacific Northwest lineages could mean complete loss of many unique alleles. At neutral markers, this loss would not be a major conservation concern, but it portends an analogous loss of diversity at evolutionarily significant loci.

The high genetic structure and uniqueness in the southern range of the snowshoe hare reflect a common phylogeographic pattern among North American species. Regional comparative studies emphasize that the Pacific Northwest and U.S. Rockies are hotspots of genetic diversity for many species (Soltis *et al.* 1997; Swenson & Howard 2005; Shafer *et al.* 2010a). Although there are few rangewide studies for boreal species, they typically corroborate the cryptic genetic distinctiveness of these southern populations in the context of the species' entire North American range (Wooding & Ward 1997; Arbogast 1999; Arbogast & Kenagy 2001; Klicka *et al.* 2011; van Els *et al.* 2012). Collectively, these findings support Hampe and Petit's (2005) call for prioritizing conservation of southern edge populations of boreal species.

For snowshoe hares and many other boreal species in North America, southern populations may already be losing genetic diversity due to anthropogenic change such as habitat fragmentation (desert bighorn sheep, *Ovis canadensis nelsoni*; Epps *et al.* 2005) and climate change (alpine chipmunk, *Tamias alpinus*; Rubidge *et al.*

2012). The range of snowshoe hares has contracted northward throughout the previous century, primarily related to habitat loss and conversion, with potential contributions from harvest and climate change (Hodges 2000; NatureServe 2014). Populations in West Virginia, North Carolina, Tennessee and Virginia have declined. Snowshoe hares are extirpated from Ohio, New Jersey and North Carolina and possibly extirpated from Maryland (NatureServe 2014). They are listed as critically imperilled (S1) in Virginia, imperilled (S2) in New Mexico and vulnerable (S3) in Pennsylvania, Utah and Nevada. In California, the subspecies *L. a. taahoensis* is a state-listed Species of Special Concern.

In the face of certain climate change with uncertain impacts, it is difficult to predict how species conservation efforts can best be prioritized to maximize long-term persistence. Although we cannot anticipate the unforeseen, we can use our understanding of the present to heed the advice of geneticist Otto Frankel (1974) that 'at this point of decision-making it may be our evolutionary responsibility to keep evolutionary options open so far as we can'. Using historical processes as a guide, an emphasis on conserving southern edge populations seems prudent for this strongly interacting prey species.

## Acknowledgements

We thank the Canadian and U.S. hunting and trapping community, agency biologists, university researchers and private citizens for donating more than 1000 snowshoe hare and black-tailed jackrabbit genetic samples for this study. We are especially grateful to N. Berg, S. Carriere, J. Ivan, H. Jolicoeur, J. MacCracken, B. McIntosh and P. Zevit for their efforts to fill in critical sampling gaps. Biologists with the U.S. Forest Service and state agencies provided invaluable help with permitting and field logistics. Special thanks to D. Wager, J. Wager, C. Brown and M. Strauser for their dedicated assistance in the field and laboratory. This work was funded by the National Science Foundation Grant 0817078, Natural Sciences and Engineering Research Council (Canada), U.S. National Park Service, and University of Montana, the Portuguese Fundação para a Ciência e a Tecnologia (FCT) and the FEDER European Social Fund (PTDC/BIA-EVF/115069/2009). J.M.-F. and PCA were funded by Portuguese Foundation for Science and Technology grants (SFRH/BPD/43264/2008 and SFRH/BSAB/1278/2012, respectively, cofunded by the European Social Fund) and Luso-American Development Foundation (FLAD).

## References

- Alves PC, Ferrand N, Suchentrunk F, Harris DJ (2003) Ancient introgression of *Lepus timidus* mtDNA into *L. granatensis* and *L. europaeus* in the Iberian Peninsula. *Molecular Phylogenetics and Evolution*, **27**, 70–80.
- Alves PC, Melo-Ferreira J, Branco M *et al.* (2008) Evidence for genetic similarity of two allopatric European hares (*Lepus*

- corsicanus* and *L. castroviejoi*) inferred from nuclear DNA sequences. *Molecular Phylogenetics and Evolution*, **46**, 1191–1197.
- Arbogast BS (1999) Mitochondrial DNA phylogeography of the new world flying squirrels (*Glaucomys*): implications for pleistocene biogeography. *Journal of Mammalogy*, **80**, 142–155.
- Arbogast BS, Kenagy GJ (2001) Comparative phylogeography as an integrative approach to historical biogeography. *Journal of Biogeography*, **28**, 819–825.
- Baker AJ, Marshall HD (1997) Mitochondrial control region sequences as tools for understanding evolution. In: *Avian Molecular Evolution and Systematics* (ed. Mindell DP), pp. 51–82. Academic Press, San Diego, California.
- Benjamini Y, Hochberg Y (1995) Controlling the false discovery rate: a practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society, Series B*, **57**, 289–300.
- Brown WM, George JM, Wilson AC (1979) Rapid evolution of animal mitochondrial DNA. *Proceedings of the National Academy of Sciences, USA*, **76**, 1967–1971.
- Burton C, Krebs CJ, Taylor EB (2002) Population genetic structure of the cyclic snowshoe hare (*Lepus americanus*) in southwestern Yukon, Canada. *Molecular Ecology*, **11**, 1689–1701.
- Coulon A, Fitzpatrick JW, Bowman R *et al.* (2008) Congruent population structure inferred from dispersal behaviour and intensive genetic surveys of the threatened Florida scrub-jay (*Aphelocoma coerulescens*). *Molecular Ecology*, **17**, 1685–1701.
- Dalquest WW (1942) Geographic variation in northwestern snowshoe hares. *Journal of Mammalogy*, **23**, 166–183.
- Darriba D, Taboada GL, Doallo R, Posada D (2012) jModelTest 2: more models, new heuristics and parallel computing. *Nature Methods*, **9**, 772.
- Daubenmire R (1975) *Floristic Plant Geography of Eastern Washington and Northern Idaho*. Brigham Young University Press, Provo, Utah.
- Drummond AJ, Suchard MA (2010) Bayesian random local clocks, or one rate to rule them all. *BMC Biology*, **8**, 114.
- Drummond AJ, Rambaut A, Shapiro B, Pybus OG (2005) Bayesian coalescent inference of past population dynamics from molecular sequences. *Molecular Biology and Evolution*, **22**, 1185–1192.
- Drummond AJ, Suchard MA, Xie D, Rambaut A (2012) Bayesian phylogenetics with BEAUti and the BEAST 1.7. *Molecular Biology and Evolution*, **29**, 1969–1973.
- Dupanloup I, Schneider S, Excoffier L (2002) A simulated annealing approach to define the genetic structure of populations. *Molecular Ecology*, **11**, 2571–2581.
- Earl DA, vonHoldt BM (2011) STRUCTURE HARVESTER: a website and program for visualizing STRUCTURE output and implementing the Evanno method. *Conservation Genetics Resources*, **4**, 359–361.
- Eckert CG, Samis KE, Loughheed SC (2008) Genetic variation across species' geographical ranges: the central-marginal hypothesis and beyond. *Molecular Ecology*, **17**, 1170–1188.
- Ehrich D, Yoccoz NG, Ims RA (2009) Multi-annual density fluctuations and habitat size enhance genetic variability in two northern voles. *Oikos*, **118**, 1441–1452.
- van Els P, Cicero C, Klicka J (2012) High latitudes and high genetic diversity: phylogeography of a widespread boreal bird, the gray jay (*Perisoreus canadensis*). *Molecular Phylogenetics and Evolution*, **63**, 456–465.
- Epps CW, Palsboll PJ, Wehausen JD *et al.* (2005) Highways block gene flow and cause a rapid decline in genetic diversity of desert bighorn sheep. *Ecology Letters*, **8**, 1029–1038.
- Evanno G, Regnaut S, Goudet J (2005) Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. *Molecular Ecology*, **14**, 2611–2620.
- Excoffier L, Laval G, Schneider S (2005) Arlequin ver. 3.0: An integrated software package for population genetics data analysis. *Evolutionary Bioinformatics Online*, **1**, 47–50.
- FAUNMAP Working Group (1994) FAUNMAP: a database documenting late Quaternary distributions of mammal species in the United States. *Illinois State Museum Scientific Papers*, **25**, 1–690.
- Frankel OH (1974) Genetic conservation: our evolutionary responsibility. *Genetics*, **48**, 53–65.
- Funk WC, McKay JK, Hohenlohe PA, Allendorf FW (2012) Harnessing genomics for delineating conservation units. *Trends in Ecology & Evolution*, **27**, 489–496.
- Gillis EA, Krebs CJ (1999) Natal dispersal of snowshoe hares during a cyclic population increase. *Journal of Mammalogy*, **80**, 933–939.
- Godbout J, Beaulieu J, Bousquet J (2010) Phylogeographic structure of jack pine (*Pinus banksiana*; Pinaceae) supports the existence of a coastal glacial refugium in northeastern North America. *American Journal of Botany*, **97**, 1903–1912.
- Gomez A, Lunt DH (2007) Refugia within refugia: patterns of phylogeographic concordance in the Iberian Peninsula. In: *Phylogeography in Southern European Refugia: Evolutionary Perspectives on the Origins and Conservation of European Biodiversity* (eds Weiss S, Ferrand N), pp. 155–188. Kluwer Academic Publishers, Dordrecht, The Netherlands.
- de Guia APO, Saitoh T (2007) The gap between the concept and definitions in the Evolutionarily Significant Unit: the need to integrate neutral genetic variation and adaptive variation. *Ecological Research*, **22**, 604–612.
- Guillot G (2008) Inference of structure in subdivided populations at low levels of genetic differentiation. The correlated allele frequencies model revisited. *Bioinformatics*, **24**, 2222–2228.
- Guillot G, Mortier F, Estoup A (2005) Geneland: a program for landscape genetics. *Molecular Ecology Notes*, **5**, 712–715.
- Hampe A, Petit RJ (2005) Conserving biodiversity under climate change: the rear edge matters. *Ecology Letters*, **8**, 461–467.
- Hansen MC, Stehman SV, Potapov PV (2010) Quantification of global gross forest cover loss. *Proceedings of the National Academy of Sciences, USA*, **107**, 8650–8655.
- Hewitt GM (1996) Some genetic consequences of ice ages, and their role in divergence and speciation. *Biological Journal of the Linnean Society*, **58**, 247–276.
- Hewitt GM (2000) The genetic legacy of the Quaternary ice ages. *Nature*, **405**, 907–913.
- Hodges KE (2000) Ecology of snowshoe hares in southern boreal and montane forests. In: *Ecology and Conservation of Lynx in the United States* (eds Ruggiero LF, Aubry KB, Buskirk SW *et al.*), pp. 163–206. University Press of Colorado, Boulder.
- IPCC (2007) *Climate Change 2007: Impacts, Adaptation, and Vulnerability*. Cambridge University Press, Cambridge, UK.
- Johnson PCD, Haydon DT (2007) Maximum-likelihood estimation of allelic dropout and false allele error rates from microsatellite genotypes in the absence of reference data. *Genetics*, **175**, 827–842.

- Kalinowski ST (2005) HP-RARE 1.0: a computer program for performing rarefaction on measures of allelic richness. *Molecular Ecology Notes*, **5**, 187–189.
- Klicka J, Spellman GM, Winker K, Chua V, Smith BT (2011) A phylogeographic and population genetic analysis of a wide-spread, sedentary North American bird: the hairy woodpecker (*Picoides villosus*). *The Auk*, **128**, 346–362.
- Korstanje R, Gillissen GF, Versteeg SA *et al.* (2003) Mapping of rabbit microsatellite markers using chromosome-specific libraries. *Journal of Heredity*, **94**, 161–169.
- Koven CD (2013) Boreal carbon loss due to poleward shift in low-carbon ecosystems. *Nature Geoscience*, **6**, 452–456.
- Krebs CJ, Boonstra R, Boutin S, Sinclair ARE (2001) What drives the 10-year cycle of snowshoe hares? *BioScience*, **51**, 25–35.
- Librado P, Rozas J (2009) DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. *Bioinformatics*, **25**, 1451–1452.
- Mantel N (1967) The detection of disease clustering and a generalized regression approach. *Cancer Research*, **27**, 209–220.
- Matthee CA, van Vuuren BJ, Bell D, Robinson TJ (2004) A molecular supermatrix of the rabbits and hares (Leporidae) allows for the identification of five intercontinental exchanges during the Miocene. *Systematic Biology*, **53**, 433–447.
- Meirmans PG (2012) The trouble with isolation by distance. *Molecular Ecology*, **21**, 2839–2846.
- Melo-Ferreira J, Boursot P, Suchentrunk F, Ferrand N, Alves PC (2005) Invasion from the cold past: extensive introgression of mountain hare (*Lepus timidus*) mitochondrial DNA into three other hare species in northern Iberia. *Molecular Ecology*, **14**, 2459–2464.
- Melo-Ferreira J, Boursot P, Randi E *et al.* (2007) The rise and fall of the mountain hare (*Lepus timidus*) during Pleistocene glaciations: expansion and retreat with hybridization in the Iberian Peninsula. *Molecular Ecology*, **16**, 605–618.
- Melo-Ferreira J, Boursot P, Carneiro M *et al.* (2012) Recurrent introgression of mitochondrial DNA among hares (*Lepus spp.*) revealed by species-tree inference and coalescent simulations. *Systematic Biology*, **61**, 367–381.
- Mills LS, Zimova M, Oyler J *et al.* (2013) Camouflage mismatch in seasonal coat color due to decreased snow duration. *Proceedings of the National Academy of Sciences, USA*, **110**, 7360–7365.
- Moore WS (1995) Inferring phylogenies from mtDNA variation: mitochondrial-gene trees versus nuclear-gene trees. *Evolution*, **49**, 718–726.
- Moritz C (1994) Defining evolutionarily significant units for conservation. *Trends in Ecology & Evolution*, **9**, 373–375.
- Moritz C, Patton JL, Conroy CJ *et al.* (2008) Impact of a century of climate change on small-mammal communities in Yosemite National Park, USA. *Science*, **322**, 261–322.
- Mougel F, Mounolou JC, Monnerot M (1997) Nine polymorphic microsatellite loci in the rabbit, *Oryctolagus cuniculus*. *Animal Genetics*, **28**, 58–71.
- NatureServe (2014) *NatureServe Explorer: An Online Encyclopedia of Life [web Application]. Version 7.1*. NatureServe, Arlington, Virginia. <http://explorer.natureserve.org>
- Nei M (1972) Genetic distance between populations. *The American Naturalist*, **106**, 283–292.
- Nei M (1978) Estimation of average heterozygosity and genetic distance from a small number of individuals. *Genetics*, **89**, 583–590.
- van Oosterhout C, Hutchinson WP, Wills DPM, Shipley P (2004) MICRO-CHECKER: software for identifying and correcting genotyping errors in microsatellite data. *Molecular Ecology Notes*, **4**, 535–538.
- Peakall R, Smouse PE (2006) GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. *Molecular Ecology Notes*, **6**, 288–295.
- Pielou EC (1991) *After the Ice Age: The Return of Life to Glaciated North America*. University of Chicago Press, Chicago, Illinois.
- Pierpaoli M, Riga F, Trocchi V, Randi E (1999) Species distinction and evolutionary relationships of the Italian hare (*Lepus corsicanus*) as described by mitochondrial DNA sequencing. *Molecular Ecology*, **8**, 1805–1817.
- Powers RP, Coops NC, Morgan JL *et al.* (2012) A remote sensing approach to biodiversity assessment and regionalization of the Canadian boreal forest. *Progress in Physical Geography*, **37**, 36–62.
- Pritchard JK, Stephens M, Donnelly P (2000) Inference of population structure using multilocus genotype data. *Genetics*, **155**, 945–959.
- Provan J, Bennett KD (2008) Phylogeographic insights into cryptic glacial refugia. *Trends in Ecology & Evolution*, **23**, 564–571.
- Raymond M, Rousset F (1995) An exact test for population differentiation. *Evolution*, **49**, 1283–1286.
- Rico C, Rico I, Webb N, Smith DB, Hewitt G (1994) Four polymorphic microsatellite loci for the European wild rabbit, *Oryctolagus cuniculus*. *Animal Genetics*, **25**, 367.
- Rousset F (1997) Genetic differentiation and estimation of gene flow from F-statistics under isolation by distance. *Genetics*, **145**, 1219–1228.
- Rousset F (2008) Genepop'007: a complete reimplementation of the Genepop software for Windows and Linux. *Molecular Ecology Resources*, **8**, 103–106.
- Rubidge EM, Patton JL, Lim M *et al.* (2012) Climate-induced range contraction drives genetic erosion in an alpine mammal. *Nature Climate Change*, **2**, 285–288.
- Ryder OA (1986) Species conservation and systematics: the dilemma of the subspecies. *Trends in Ecology & Evolution*, **1**, 9–10.
- Schwartz MK, Mills LM, Ortega Y, Ruggiero LF, Allendorf FW (2003) Landscape location affects genetic variation of Canada lynx (*Lynx canadensis*). *Molecular Ecology*, **12**, 1807–1816.
- Schwartz MK, Pilgrim KL, McKelvey KS, Rivera PT, Ruggiero LF (2007) DNA markers for identifying individual snowshoe hares using field-collected pellets. *Northwest Science*, **81**, 316–322.
- Shafer ABA, Cote SD, Coltman DW (2010a) Hot spots of genetic diversity descended from multiple Pleistocene refugia in an alpine ungulate. *Evolution*, **65**, 125–138.
- Shafer ABA, Cullingham CI, Cote SD, Coltman DW (2010b) Of glaciers and refugia: a decade of study sheds new light on the phylogeography of northwestern North America. *Molecular Ecology*, **19**, 4589–4621.
- Shugart HH, Leemans R, Bonan GB (2005) *A Systems Analysis of the Global Boreal Forest*, pp. 565. Cambridge University Press, Cambridge.
- Soltis DE, Gitzendanner MA, Strenge DD, Soltis PS (1997) Chloroplast DNA intraspecific phylogeography of plants from the Pacific Northwest of North America. *Plant Systematics and Evolution*, **206**, 353–373.

- Soltis DE, Morris AB, McLachlan JS, Manos PS, Soltis PS (2006) Comparative phylogeography of unglaciated eastern North America. *Molecular Ecology*, **15**, 4261–4293.
- Strimmer K (2008) fdrtool: a versatile R package for estimating local and tail area-based false discovery rates. *Bioinformatics*, **24**, 1461–1462.
- SurrIDGE AK, Bell DJ, Rico C, Hewitt GM (1997) Polymorphic microsatellite loci in the European rabbit (*Oryctolagus cuniculus*) are also amplified in other lagomorph species. *Animal Genetics*, **28**, 302–305.
- Swenson NG, Howard DJ (2005) Clustering of contact zones, hybrid zones, and phylogeographic breaks in North America. *American Naturalist*, **166**, 581–591.
- Tracy LN, Jamieson IG (2011) Historic DNA reveals contemporary population structure results from anthropogenic effects, not pre-fragmentation patterns. *Conservation Genetics*, **12**, 517–526.
- Vigilant L, Stoneking M, Harpending H, Hawkes K, Wilson AC (1991) African populations and the evolution of human mitochondrial DNA. *Science*, **253**, 1503–1507.
- Vogler AP, DeSalle R (1994) Diagnosing units of conservation management. *Conservation Biology*, **8**, 354–363.
- Weir BS, Cockerham CC (1984) Estimating F-statistics for the analysis population structure. *Evolution*, **38**, 1358–1370.
- Williams MAJ, Dunkerley DL, De DP, Kershaw AP, Stokes T (1993) *Quaternary Environments*. Edward Arnold, New York.
- Wooding S, Ward R (1997) Phylogeography and Pleistocene evolution in the North American black bear. *Molecular Biology and Evolution*, **14**, 1096–1105.

---

E.C., L.S.M. and K.E.H. designed the research; E.C. conducted field sampling and laboratory work; L.S.M. and K.E.H. contributed some hare genetic samples; P.C.A. and J.M.F. provided laboratory training and assistance; E.C. and J.M.F. analysed data and wrote this work with direction, assistance and editorial review from L.S.M., K.E.H. and P.C.A.

---

### Data accessibility

All Cytb and CR sequences from this project have been deposited in GenBank (Accession nos KF781351–KF781437; KF804153–KF805042; HM771306–HM771308).

Sample details, sequence alignments, microsatellite genotypes, and input files have been deposited in the Dryad Digital Repository (Conservation implications of the evolutionary history and genetic diversity hotspots of the snowshoe hare. <http://dx.doi.org/10.5061/dryad.dh63p>).

### Supporting information

Additional supporting information may be found in the online version of this article.

**Appendix S1** Sources of genetic samples analysed in this study.

**Appendix S2** Microsatellite loci diversity.

**Appendix S3** DNA extraction and genotyping methods.

**Appendix S4** STRUCTURE hierarchical analysis results under a model of admixture and uncorrelated allele frequencies.

**Appendix S5** Analysis of the Boreal cluster in the second round of STRUCTURE hierarchical analysis.

**Appendix S6** Highest mean posterior density Geneland results for a model with uncorrelated allele frequencies and null alleles filtered.

**Appendix S7** Pairwise  $F_{ST}$  and Nei's D calculated across eight microsatellite loci.

**Appendix S8** Population allelic richness plotted against latitude and longitude.

**Appendix S9** Mitochondrial control region median-joining network.

**Appendix S10** Mitochondrial control region pairwise  $F_{ST}$ .

**Appendix S11** Mitochondrial control region diversity statistics.

**Appendix S12** Haplotype diversity plotted against latitude and longitude.

**Appendix S13** Nucleotide diversity plotted against latitude and longitude.